Using eDNA (single species qPCR assays) for aquatic invasive species in AK





Early Detection/ Rapid Response/Treatment Efficacy

Case study on Kenai Peninsula



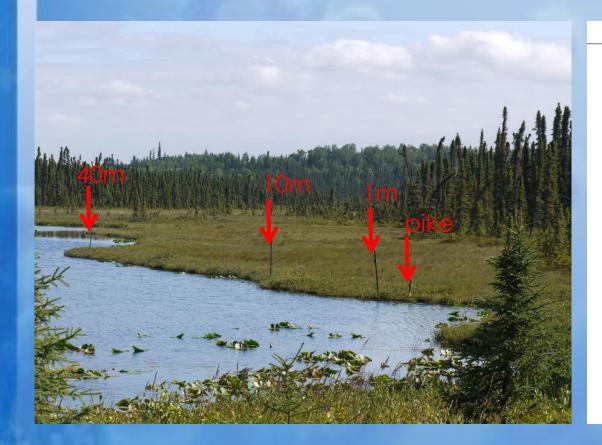
Potential of Environmental DNA to Evaluate Northern Pike (*Esox lucius*) Eradication Efforts: An Experimental Test and Case Study

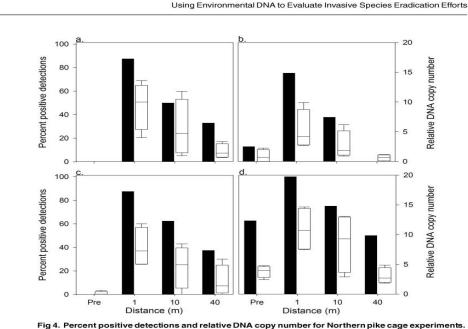
Kristine J. Dunker¹, Adam J. Sepulveda²*, Robert L. Massengili³, Jeffrey B. Olsen⁴, Ora L. Russ⁴, John K. Wenburg⁴, Anton Antonovich¹



N. pike case study: take home 1

Detection probability decreases with distance*





Percent positive detections (filled bars) and relative DNA copy number (unfilled, box plots) for Northern pike cage experiments in Denise (a), Gensle (b), Little Bear (c) and Tiny (d) Lakes near Soldotna, AK. For the box plots, the dark horizontal line represents the mean, with the box representing the 25th and 75th percentiles and the whiskers the 5th and 95th percentiles. Water samples (n = 8 per lake treatment) were analyzed for Northern pike DNA prior (Pre) to introduction of caged Northern pike and then 7 days after

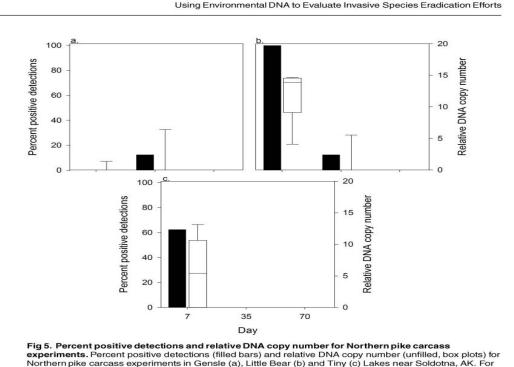
introductions at 1 m, 10 m, and 40 m away from each cage.

doi:10.1371/journal.pone.0162277.g004

N. pike case study: take home 2

Detection probability decreases with time*





the box plots, the dark horizontal line represents the mean, with the box representing the 25th and 75th percentiles and the whiskers the 5th and 95th percentiles. Water samples (n = 8 per lake per day) were

analyzed for Northern pike DNA 7, 35 and 70 days after carcass additions.

N. pike case study: take home 3

DNA, together with other methods, confirms eradication*



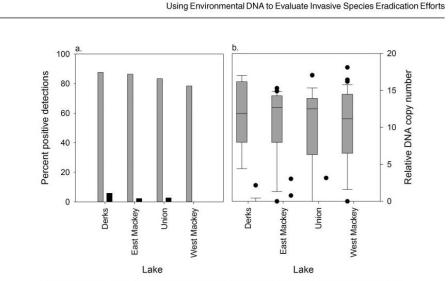
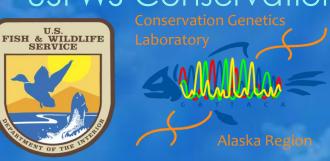


Fig 6. Percent positive detections and relative DNA copy number before and after rotenone eradication treatments. (a) Percent positive detections and (b) relative DNA copy number before (gray filled) and after (black filled) rotenone eradication treatments in Derks, East Mackey, Union and West Mackey lakes near Soldonta, AK. Relative DNA copy numbers are displayed as box plots, with the dark horizontal line representing the mean, the box representing the 25th and 75th percentiles, the whiskers representing the 5th and 95th percentiles and the filled circles representing outliers. eDNA water samples were collected ~ 30 days before and ~ 230 days after the rotenone treatments.

doi:10.1371/journal.pone.0162277.g006

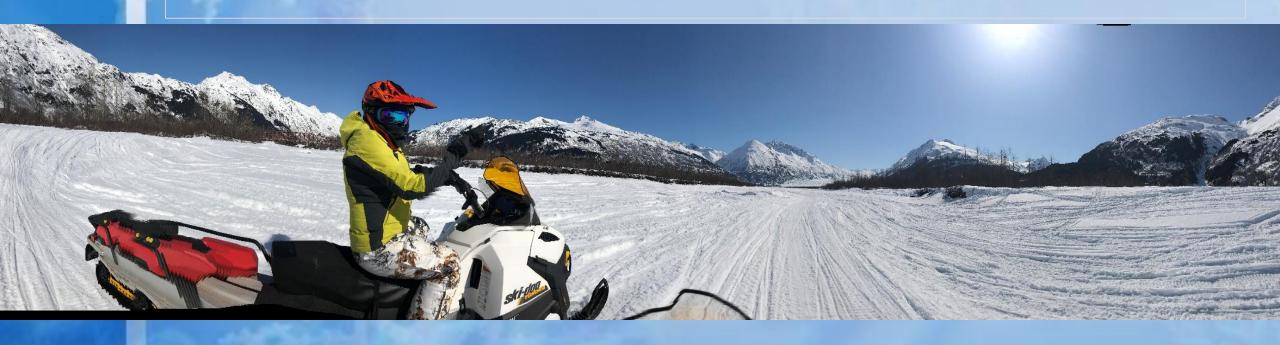
Seasonal Variation in the Detection of Northern Pike eDNA in a Southcentral Alaska Lake

Ora Russ – USFWS Conservation Genetics Lab Catherine Bradley – USFWS Fairbanks FWFO Jeffrey Olsen – USFWS Conservation Genetics Lab Jason Everett – USFWS Conservation Genetics Lab John Wenburg – USFWS Conservation Genetics Lab



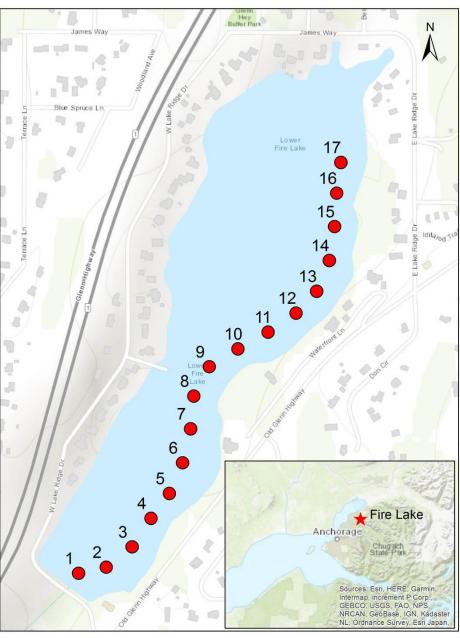
Objective:

Determine if there is a difference in detection probability of Northern Pike eDNA (in a known Northern Pike lake) among seasons.



Challenge for managers...
accessibility to remote locations

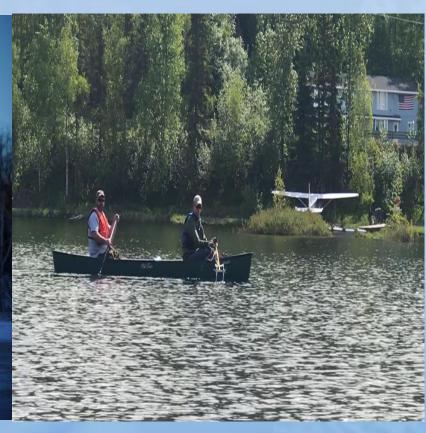
Fire Lake / 17 sites



Methods: field (1L water samples)







Fall 2016

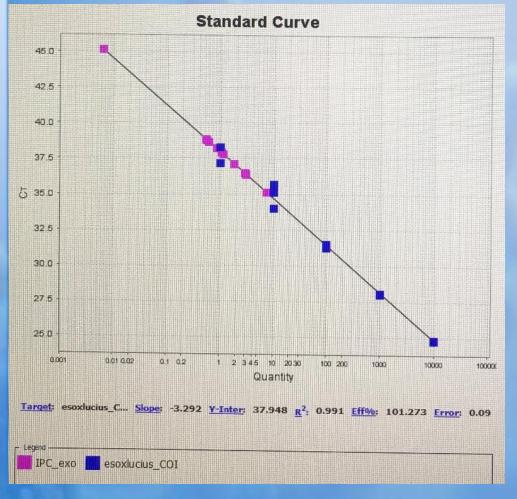
Winter 2018

Summer 2018

- none sampling occasion (day) each season
- * sampling day assumed to be representative of the season

Methods: Lab & Statistical

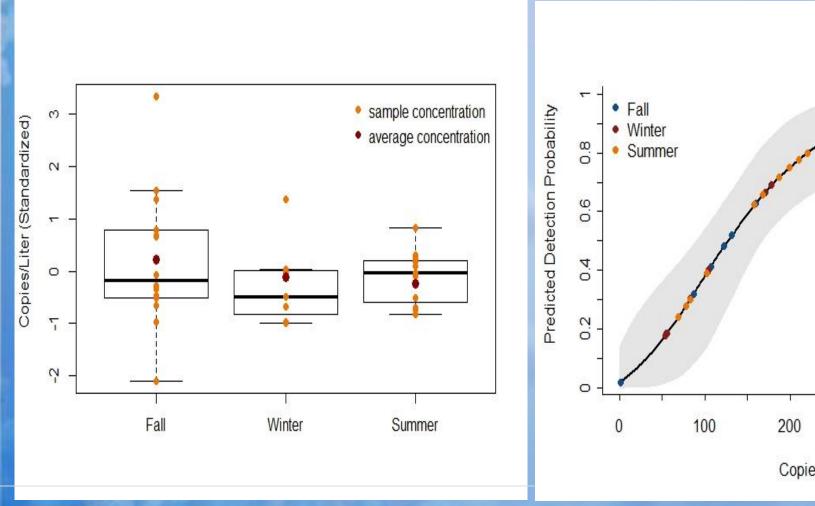
Single species qPCR assay (Northern Pike COI)

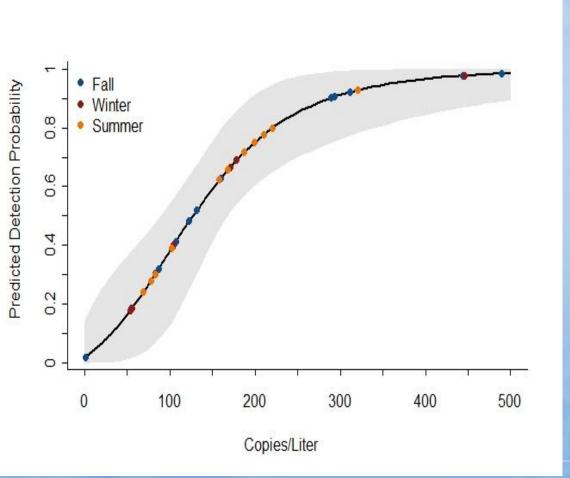


Estimate relative eDNA concentration (copies/L) for each of 3 "Seasons" and put that data into Occupancy model...

Compare eDNA detection probability using a hierarchical Bayesian framework (following Kery and Schaub 2012)

Results:





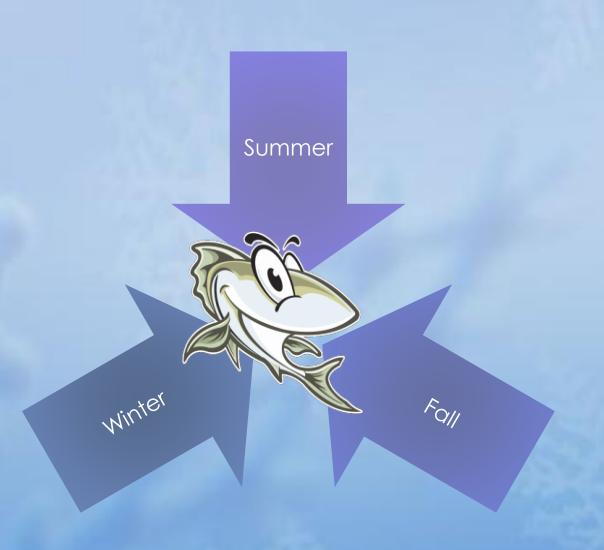
^{*}Estimated copies/L varied, but were not significantly different among seasons (Kruskal-Wallis chi-squared = 1.08, p-value = 0.58)

^{*}eDNA concentration strongly influences eDNA detection probability: 204 copies/L is avg eDNA conc. @which detection probability is 66.9%.

Discussion:

Northern Pike eDNA detection probability is driven by eDNA concentration (copies/Liter) more so than season.

★ More sampling sites or replication of samples may be required in Winter to achieve the same detection probability as Fall and Summer.



Conclusions:

Are you eDNA ready?....

- Sampling strategy is key!! (whey we explored seasonal?)
- ★Pilot studies essential to giving a good baseline and high confidence in eDNA results

★eDNA can be a great complementary tool to observational data. Not meant to replace traditional tools. (Kenai pike study used netting + eDNA)



Elodea....From a cute little aquarium plant to a cold water adapted AK invasive plant



How should *Elodea* eDNA be applied moving forward? -A controlled field study

USFWS: Jimmy Fox, Anna-Marie Benson, Ora Russ



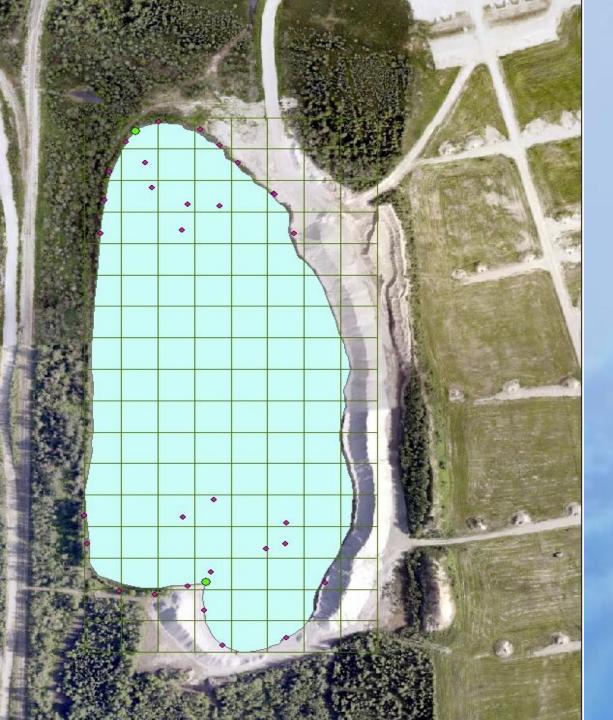
DOD Natural Resources Ft. Wainright: 5.6 hectacre pond

eDNA assays for elodea

- > USACE has developed three qPCR assays
 - Generic elodea, E. canadensis, E. nuttallii

Objective: Detection probability using occupancy modeling

- 1) 95% confident of detecting 80% probability of site occupancy of *Elodea* eDNA (50 m, 75m, 100m) Limit of Dectection
- 2) Estimate how quickly *Elodea* eDNA can be detected in water samples at the above distances (one month, 3-4 months, 6-8 months) after introduction.







Elodea was introduced to pond in custom mesh screen buckets (EloCondos) on August 14, 2018

Location of captive Elodea plants (green dots) and water samples (pink dots) collected at the Small Arms Complex Pond (SACpond) on 26 September 2018 on Fort Wainwright, Alaska. The 25m x 25m grid was used to define our sample units.

- *No detections from Sept 2018 sampling (N=30 replicated x 2)
- *March 26, 2019 under ice sampling (lab results in process)
- *Summer 2019 sampling planned

Happy to take questions.....

